

UMass Soil & Plant Nutrient Testing Laboratory 203 Paige Laboratory

203 Paige Laboratory
161 Holdsworth Way
Amherst, MA 01003
413-545-2311
<u>soiltest@umass.edu</u>
http://soiltest.umass.edu/

USE THIS FORM FOR PLANT NUTRIENT SAMPLE SUBMISSION FOR VEGETABLE CROPS. (See page 2 for sampling instructions, fees, and description of services.)

Main Contact:	Send Copy to:	Method of receiving results
Name:	Name:	US Mail (Please include
Business Name:	Business Name:	\$2 per order for postage & handling)
Street Address:	Street Address:	
City, State, Zip:	City, State, Zip:	Email
Phone:	Phone:	Send copies to:
Email Address:	Email Address:	

	LAB#	Sample ID	Test Requested
(Leav	ave blank)	(You create this)	Standard (\$45) or Standard w/o Nitrogen (\$30)

	Order Total \$		
Sample Information	Complete this section for problem diagnosis		
Crop, management, and soil information			
Date Sampled:	If leaves are discolored, does color variation occur:		
Crop: Variety:	Along leaf margins Interveinal In spots Over entire leaf		
Growth Stage:			
Plant spacing or population:	Leaves first affected at shoot: tip base over entire shoot		
Lime: tons/Acre applied on: (date)			
Fertilizer rate(s) and date(s):	Symptoms first seen:(month & growth stage)		
	Describe additional symptoms:		
Soil series (if known):			

Office U	Jse Only
Received	Due
Check#	PO#
Cash	Date

Please make check payable to the University of Massachusetts or "UMass"

General Sampling Procedure:

For a routine evaluation of plant status, we compare nutrient levels to data collected in scientific literature. It is extremely important to collect samples at the growth stage and from the plant part for which plant nutrient data is available.

Specific sampling instructions for the most common commercially grown vegetables in New England are provided here. This is not a complete list. **Contact the lab for crops not listed here to be sure plant nutrient data is available and for sampling instructions.**

Samples should reflect areas with uniform management and soil type. Where differences occur within a block, sampling should be refined to represent these changes. Samples should represent only one cultivar, and should be collected from several different plants within the block.

When you suspect a nutrient deficiency, always attempt to collect one sample from plants in the affected area and a second sample from plants of the same variety in an area showing normal growth. This will allow for direct comparison of nutrient levels and may aid in diagnosing specific nutrient deficiencies.

When collecting tissue samples, avoid diseased or dead plant material, tissue damaged by equipment or insects, and plant tissue stressed by excessive heat, cold, or moisture. Do not sample seed because it does not reflect the nutrient status of the whole plant.

After collecting your composite sample, it is a good idea to rinse the tissue with clean water to remove pesticides, foliar applied nutrients, and soil particles. Place wet samples on a clean paper towel to air dry. Once dry, carefully place sample in a **small <u>paper</u> bag labeled with your sample ID** and complete the submission form. Hand deliver or mail the sample, submission form, and check or money order payable to UMass to the address listed on the front of this form.

Plant Tissue Nutrient Test Descriptions & Fees

Standard Tissue Nutrient Test: \$45.00

A determination of the Total Tissue P, K, Ca, Mg, Zn, Cu, Mn, Fe, and B. Analysis by ICP Spectroscopy of acid wet digestion in Nitric Acid, Hydrochloric Acid, and Hydrogen Peroxide in a block digester. Also included, Total Nitrogen by catalytic combustion.

Standard Tissue Test Without Total Nitrogen: \$30.00 Same as standard tissue test but without Total Nitrogen

<u>Crop – Plant part collected – Growth Stage</u>

Beans – 10-15 uppermost recent fully-developed trifoliate leaves – Summer Beets –20-25 mature leaves from new growth – 4-6 weeks after seeding OR 8-10
weeks after seeding
Broccoli or Cauliflower – 12-15 mature leaves from new growth – At heading
Brussels Sprouts – 12-15 mature leaves from new growth– Maturity
Cabbage – 15-20 whole tops – 2-6 weeks old
Cabbage – 12-15 wrapper leaves – 2-3 months old Cabbage – 15-20 midribs from wrapper leaves – Mature plants
Cantaloupe or Muskmelon – 12 unfurled leaves (5th leaf from tip) – Flower start
to small fruit OR Small fruit to harvest
Carrots – 15 mature leaves from new growth -Middle of growing season
Carrots – 15-20 oldest leaves – Mature plants Celery, Field – 12-15 petioles
from most recent fully- developed leaves – 6 week old plants
Celery, Field – 12-15 mature leaves from new growth – Mature plants, non-
flowering
Celery, Greenhouse – 12-15 mature leaves from new growth – 6 weeks after
transplanting
Collards or Kale – 12-15 mature leaves from new growth – Middle of growing
season
Corn, Sweet – 10-15 fully mature leaves from below the whorl – Prior to
tasselling
Corn, Sweet – 10-15 entire leaves at the ear node – At tasselling
Cucumber – 12 leaf blades (5th leaf from tip) – Flower start to small fruit OR Small
fruit to harvest
Potato – 25-30 most recent fully-developed leaves – Plants 30 cm tall OR Tubers ½ grown
Pumpkin – 15-20 mature leaves from new growth – Middle of growing
season
Radish – 30-35 most recent fully developed leaves – Middle of growing
season
Spinach, Field – 15-20 most recent fully developed leaves
– 25-30 days old OR Mature plants
Summer Squash – 12 blades from most recent fully- developed leaves –
Summer
Tomato – 15-20 compound leaves adjacent to top influor- escences - Mid-bloom
Watermelon – 10-12 unfurled leaves – Flower start to small fruit
Watermelon – 12-15 mature leaves from new growth – Mature plant, small
fruit stage
Watermelon – 12-15 unfurled leaves (5th leaf from tip) – Older fruit to harvest
Zucchini – 12-15 mature leaves from new growth – Mature plants, non-fruiting